ZooKeys 852: 1–21 (2019)
doi: 10.3897/zookeys.852.28964
http://zookeys.pensoft.net

RESEARCH ARTICLE

A peer-reviewed open-access journal
ZooKeys
Launched to accelerate biodiversity research

Two new species of *Acanthobothrium* Blanchard, 1848 (Onchobothriidae) in *Narcine entemedor* Jordan & Starks, 1895 (Narcinidae) from Acapulco, Guerrero, Mexico

Francisco Zaragoza-Tapia[1], Griselda Pulido-Flores[1], Juan Violante-González[2], Scott Monks[1]

1 *Universidad Autónoma del Estado de Hidalgo, Centro de Investigaciones Biológicas, Apartado Postal 1-10, C.P. 42001, Pachuca, Hidalgo, México* **2** *Universidad Autónoma de Guerrero, Unidad Académica de Ecología Marina, Gran Vía Tropical No. 20, Fraccionamiento Las Playas, C.P. 39390, Acapulco, Guerrero, México*

Corresponding author: *Scott Monks* (monks.scott@gmail.com)

Academic editor: *B. Georgiev* | Received 14 August 2018 | Accepted 25 April 2019 | Published 5 June 2019

http://zoobank.org/0CAC34BD-1C75-415F-973D-C37E4557D06F

Citation: Zaragoza-Tapia F, Pulido-Flores G, Violante-González J, Monks S (2019) Two new species of *Acanthobothrium* Blanchard, 1848 (Onchobothriidae) in *Narcine entemedor* Jordan & Starks, 1895 (Narcinidae) from Acapulco, Guerrero, Mexico. ZooKeys 852: 1–21. https://doi.org/10.3897/zookeys.852.28964

Abstract

Two species of *Acanthobothrium* (Onchoproteocephalidea: Onchobothriidae) are described from the spiral intestine of *Narcine entemedor* Jordan & Starks, 1895, in Bahía de Acapulco, Acapulco, Guerrero, Mexico. Based on the four criteria used for the identification of species of *Acanthobothrium*, *A. soniae* **sp. nov.** is a Category 2 species (less than 15 mm in total length with less than 50 proglottids, less than 80 testes, and with the ovary asymmetrical in shape). *Acanthobothrium vidali* **sp. nov.** is a Category 6 species (more than 15 mm in total length with more than 50 proglottids, fewer than 80 testes, and the ovary is asymmetrical). The new species differ from similar species from the Pacific Ocean by total length, the number of proglottids, diameter of the accessory sucker, the length of the cirrus sac, the number of testes per proglottid and the measurements of hooks. With the recognition of *A. soniae* **sp. nov.** and *A. vidali* **sp. nov.**, 42 species of *Acanthobothrium* have been reported from the Pacific coast of the Americas. This is the first report of species of *Acanthobothrium* from a member of *Narcine* from Mexico and it brings the number of species reported from elasmobranchs from the Pacific Coast of Mexico to 13.

Keywords

Acanthobothrium, Elasmobranchii, Helminth, *Narcine entemedor*, Onchoproteocephalidea, Torpediniformes

Copyright *Francisco Zaragoza-Tapia et al.* This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

Acanthobothrium Blanchard, 1848 is one of the richest genera within Onchoproteocephalidea (Maleki et al. 2013; Caira et al. 2017), but relatively few occurrences have been documented in Mexico. To date, the best-studied locality is the Gulf of California (Sea of Cortez; Mar de Cortés, Golfo de California) with the descriptions of 10 species (Appy and Dailey 1973; Caira and Burge 2001; Caira and Zahner 2001; Ghoshroy and Caira 2001). The only other species known from the Pacific Coast of Mexico was described from the more southern state of Jalisco (Monks et al. 1996) (see Merlo-Serna and García-Prieto 2016). More recently, *A. cartagenensis* Brooks & Mayes, 1980 was reported from Quintana Roo, Mexico (Caribbean) (Monks et al. 2015a) and *A. marquesi* was described from Campeche, Mexico (Gulf of Mexico) by Rodríguez-Ibarra et al. (2018). As part of a collaborative project to extend the knowledge of the helminth fauna of marine fishes in Mexico, rays were collected from the coastal waters off Acapulco, Guerrero, a region with few studies of the parasites of rays. There are six reports of parasites of rays from Acapulco, none for *Acanthobothrium* (see Merlo-Serna and García-Prieto 2016). In this paper, two new species of *Acanthobothrium* are described from *Narcine entemedor* Jordan & Starks, 1895 (Elasmobranchii: Torpediniformes: Narcinidae); one Category 2 species (Ghoshroy and Caira 2001) and one Category 6 species. The new species constitute the first records from the Southern Pacific Coast of Mexico and the first record of species of *Acanthobothrium* reported in *Narcine entemedor* from Mexico.

Materials and methods

Eleven recently killed specimens of *Narcine entemedor* Jordan & Starks, 1895 (Giant electric ray or Cortez Numbfish) were purchased from local fishermen at Playa Las Hamacas, Bahía de Acapulco, Guerrero (16°51'10.80"N, 99°53'59.02"W) in February, April, May, June 2011 and June 2012. Rays were transported to the laboratory (Universidad Autónoma de Guerrero, Unidad Académica de Ecología Marina) in ice chests containing ice where the spiral intestine was removed and opened by longitudinal incision. Collection, preservation and mounting follow Monks et al. (2015b). Stained specimens were examined using a compound photomicroscope (Leica DM-LB2) equipped with both normal light optics and differential interference contrast (DIC-Nomarski) optics. Figures were drawn with the aid of a drawing tube. Measurements are presented as ranges and number of specimens from which the measurements were taken is given in parentheses. Hook measurements follow Euzet (1959) as modified by Ghoshroy and Caira (2001). Measurements are in micrometers unless otherwise stated. Other hook terminology follows that of Caira (1985). Designation of proglottid apolysis follows Caira et al. (1999) and Franzese and Ivanov (2018). Microtriches on the scolex and peduncle were not examined using SEM, so they are referred to by the general name "microtriches" without qualifying them as proposed by

Chervy (2009). The categorical method suggested by Ghoshroy and Caira (2001) and Fyler and Caira 2006 was used to facilitate comparisons among species of *Acanthobothrium* from the Eastern Pacific Ocean and other congeners with similar morphological characters described in the Pacific Ocean. Specimens from several museums were examined, the acronyms are as follows:

CNHE (Colección Nacional de Helmintos del Instituto de Biología, Universidad Nacional Autónoma de México, México);
HWML (University of Nebraska State Museum, Harold W. Manter Laboratory, Division of Parasitology, Lincoln, Nebraska, USA);
CHE (Colección de Helmintos, Centro de Investigaciones Biológicas, Universidad Autónoma del Estado de Hidalgo, Pachuca, México).

Type material was deposited in CNHE (holotype and paratypes), HWML (paratypes), and CHE (paratypes). Application and validity of scientific names, authorities, and common names of fish are in accord with Froese and Pauly (2018) and Last et al. (2016).

Systematic accounts

Order Onchoproteocephalidea Caira, Jensen, Waeschenbach, Olson & Littlewood, 2014
Family Onchobothriidae Braun, 1900
Genus *Acanthobothrium* Blanchard, 1848
Type species *Acanthobothrium coronatum* (Rudolphi, 1819) Blanchard, 1848

***Acanthobothrium soniae* sp. nov.**
http://zoobank.org/4E0635D5-3E38-4FEC-8192-4B0AEF53C76C
Figures 1A–E; 2A–D

Type material. Holotype (CNHE-11136), 3 paratypes (CNHE-11137), 3 paratypes (HWML-139978), and 1 paratype (CHE-P00081).

Other material examined. *Acanthobothrium bullardi* Ghoshroy & Caira, 2001 (CNHE–4046, México) paratype; *A. campbelli* Marques, Brooks & Monks, 1995 (CNHE–3033, Costa Rica; HWML–38546, Costa Rica) voucher and paratype; *A. costarricense* Marques, Brooks & Monks, 1995 (CNHE–3034, Costa Rica) 2 vouchers; *A. dasi* Ghoshroy & Caira, 2001 (CNHE–4044, México; HWML–15549, 15550, 15551, México) 4 paratypes; *A. franus* Marques, Centritto & Stewart, 1997 (CNHE–3140, Costa Rica) paratype; *A. inbiorium* Marques, Centritto & Stewart, 1997 (CNHE–3138, Costa Rica) paratype; *A. puntarenasense* Marques, Brooks & Monks, 1995 (CNHE–4176, Costa Rica) paratype; *A. rajivi* Ghoshroy & Caira, 2001 (CNHE–4039, México) paratype; *A. vargasi* Marques, Brooks & Monks, 1995 (HWML 38545, Costa Rica).

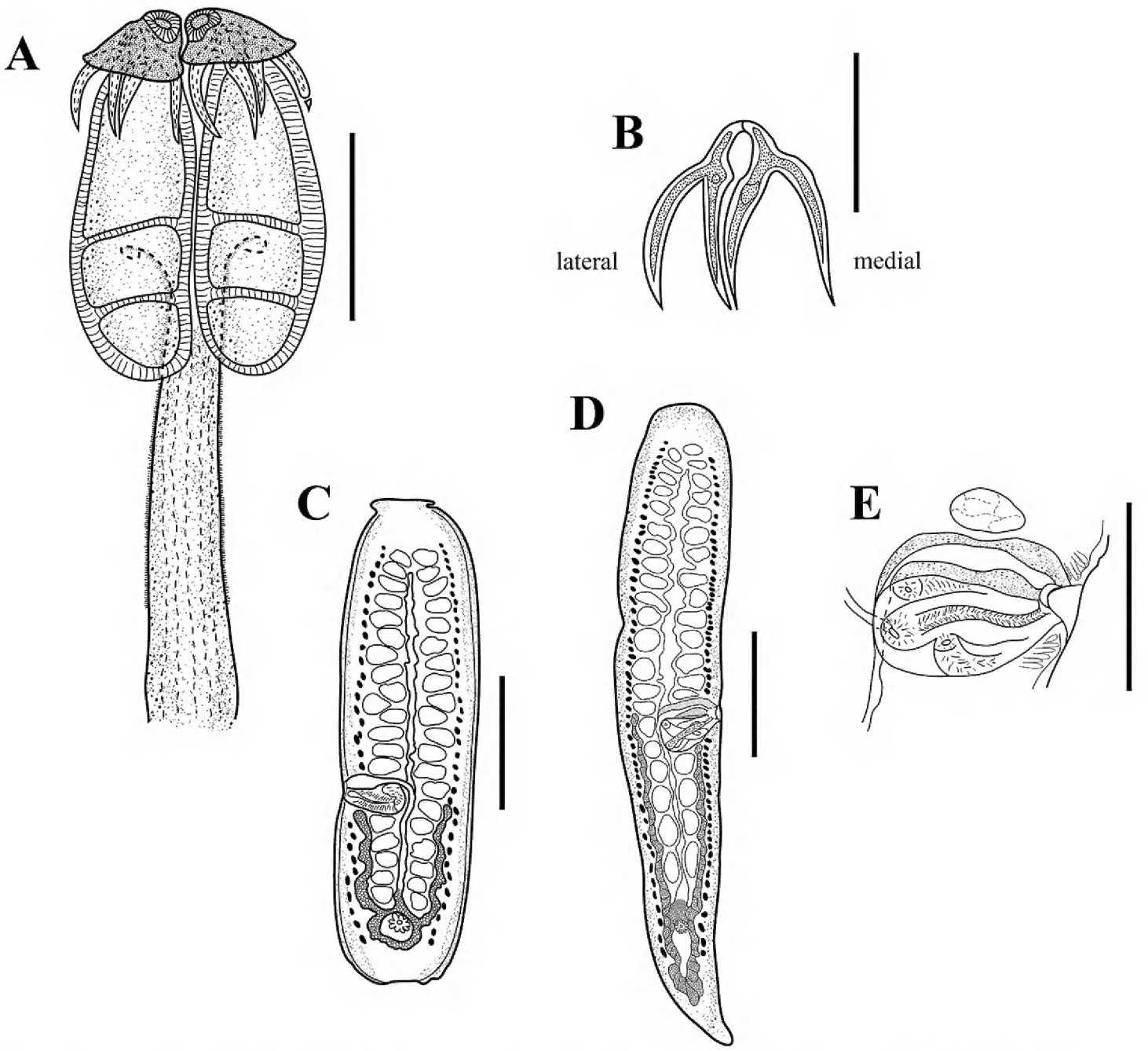

Figure 1. Holotype of *Acanthobothrium soniae* sp. nov. (CNHE-11136). **A** Scolex **B** hooks **C** mature proglottid **D** terminal proglottid **E** genitalia. Scale bars: 200 μm (**A, D**); 100 μm (**B, E**); 150 μm (**C**).

Type host. *Narcine entemedor* Jordan & Starks, 1895 (Elasmobranchii: Torpediniformes: Narcinidae).

Type locality. Bahía de Acapulco (Playa Las Hamacas: 16°51'11"N, 99°53'59"W), Guerrero, México.

Site of infection. Spiral intestine.

Quantitative descriptors of parasite populations (Bush et al. 1997). Prevalence= 9.0% (1 ray of 11 was infected); abundance = 0.73 (8 helminths in 11 rays); mean intensity = 8 (8 helminths in 1 infected ray).

Etymology. The species is named in remembrance of Sonia Virginia Flores León, former player of the Pumas Club Women's Basketball Team, UNAM, daughter of Virginia León-Règagnon and Martín Ignacio Flores-Carbajal and dear friend of SM and GP-F; she will not be forgotten.

Diagnosis. *Acanthobothrium soniae* sp. nov. is a Category 2 species. It is small, with a range of 10–13 acraspedote proglottids. The testes are wider than long with a range of

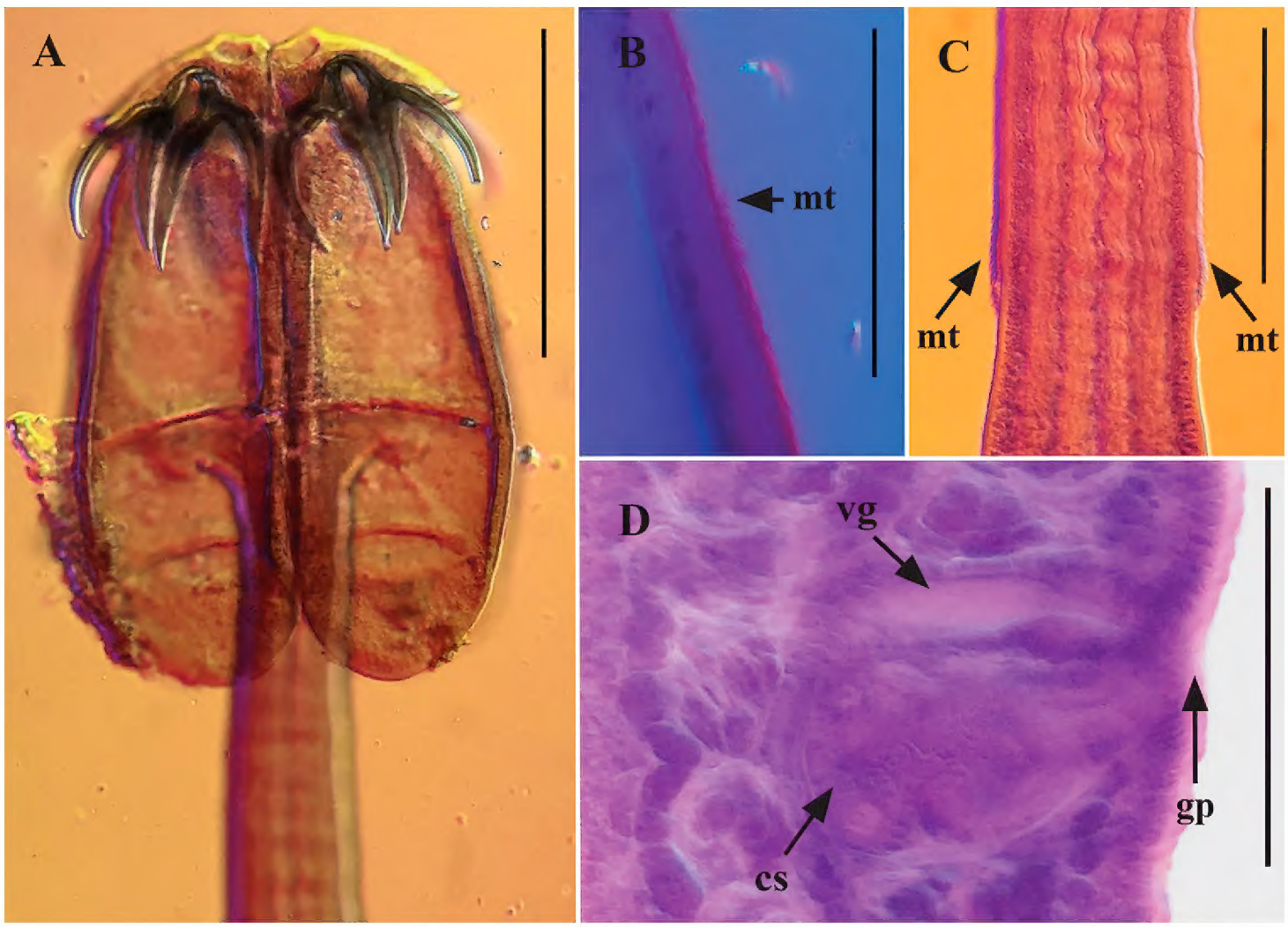

Figure 2. Light microscope photographs of holotype of *Acanthobothrium soniae* sp. nov. (CNHE-11136). **A** Scolex **B** details of bothridium **C** peduncle cephalic **D** genitalia. Scale bars: 200 μm (**A**); 40 μm (**B**), 100 μm (**C, D**). Abbreviations: mt microtriches; vg vagina; cs cirrus sac; gp genital pore.

31–47 testes per proglottid. The arms of the ovary are unequal (asymmetrical). Finally, this species also can be distinguished from similar congeners by total length, number of proglottids, diameter of accessory sucker, the length of the cirrus sac, number of testes per proglottid, and size of the hooks.

Description. [Based on 5 complete worms and 3 partial specimens] Worms 2.9–6.7 mm (n = 5) long, euapolytic; 10–13 (n = 5) proglottids per worm. Scolex 380–420 (n = 6) long by 280–320 (n = 6) wide, with four bothridia. Maximum width of scolex at level of middle loculus (Figs 1A, 2A). Bothridia free posteriorly, tri-locular, 340–380 (n = 6) long by 140–160 (n = 6) wide, with anterior muscular pad (Figs 1A, 2A). Muscular pad 105–130 (n = 6) wide, with apical sucker 45–50 (n = 6) and one pair of bipronged hooks at posterior margin (Figs 1A, 2A). Anterior loculus of bothridia 175–205 (n = 6) long; middle loculus 60–90 (n = 6) long; posterior loculus 80–95 (n = 6) long (Figs 1A, 2A); loculus length ratio (anterior:middle:posterior) 1:0.38:0.46. Velum between medial margins of bothridia in dorsal or ventral pairs not seen (Figs 1A, 2A). Hooks bipronged, hollow, with tubercle on proximal surface of axial prong; internal channels of axial and abaxial prongs continuous, smooth, the base and anterior part of each hook embedded in musculature of scolex, tips of prongs free (Figs 1A, B, 2A). Bases (handles) of medial and lateral hooks articulate to one another (Figs 1B,

2A). Lateral hook measurements (n = 6): A 43–45, B 88–105, C 83–93, D 125–138; Medial hook measurements (n = 6): A' 38–45, B' 83–108, C' 80–98, D' 125–143. Cephalic peduncle 450–630 (n = 6) long by 70–95 (n = 6) wide, not all the cephalic peduncle is covered with prominent microtriches (Figs 1A, 2C). Scolex is covered with microtriches (Fig. 2B). Proglottids acraspedote. Immature proglottids 55–110 (n = 8) long by 60–115 (n = 8) wide, mature proglottids 225–800 (n = 8) long by 125–215 (n = 8) wide (Fig. 1C), terminal proglottids 585–1,425 (n = 7) long by 160–275 (n = 7) wide (Fig. 1D). Genital pore marginal, irregularly alternating, 56%–68% (n = 7) from anterior end of proglottid; genital atrium present (Fig. 1E). Testes in single layer, arranged in two irregular columns, one on each side of the uterus (Fig. 1C, D). Testes generally wider than long in mature proglottids, 25–63 (n = 8) long by 13–28 (n = 8) wide (Fig. 1C). Total number of testes 31–47 (n = 8), aporal 16–26 (n = 8), poral 15–21 (n = 8), preporal 11–17 (n = 8), and postporal 3–5 (n = 8); all testes located anterior to ovarian isthmus. Cirrus sac pyriform, extending anteriorly (Figs 1C, D, E, 2D), 55–90 (n = 6) long by 63–96 (n = 6) wide in mature proglottids, 85–140 (n = 6) long by 48–90 (n = 6) wide in termial proglottids. Cirrus armed.

Vagina extending laterally from common genital atrium, following anterior margin of cirrus sac, weakly sinuous posteriorly along medial line of proglottid to oötype (Figs 1C, D, E, 2D); vaginal sphincter absent. Seminal receptacle not seen. Ovary inverted A-shaped in frontal view in mature and terminal proglottids (Fig. 1C, D). Arms of ovary unequal (Fig. 1C, D); aporal arm always longer than poral arm. Aporal arm 78–275 (n = 7) long in mature proglottids, 243–625 (n = 7) long in terminal proglottids; poral arm 68–213 (n = 7) long in mature proglottids, 190–550 (n = 7) long in terminal proglottids and Mehlis' gland posterior to ovarian isthmus. Vitellarium follicular form lateral bands, extending from near anterior margin of proglottid to near posterior margin of proglottid (Fig. 1C, D); follicles 15–23 (n = 6) long by 10–13 (n = 6). Uterus thick-walled, saccate, extending from anterior margin of proglottid to near posterior margin of proglottid. Excretory ducts laterally. Gravid proglottids and eggs not seen.

Remarks. There are 42 Category 2 species (sensu Ghoshroy and Caira 2001) of *Acanthobothrium* that have been described worldwide. Of these, 17 species have been found in the Pacific Ocean, 14 of which are amphi-American (Table 1).

Acanthobothrium soniae sp. nov. is a Category 2 species (sensu Ghoshroy and Caira 2001): Category 2 species have a total length ≤ 15 mm (the length of *A. soniae* sp. nov. is 2.9–6.7 mm), a strobila made up of ≤ 50 proglottids (*A. soniae* sp. nov. has 10–13 proglottids), the number of testes per proglottids ≤ 80 (*A. soniae* sp. nov. has 31–47 testes per proglottid), and the arms of the ovary are asymmetrical.

As presented in Table 1, the new species can be distinguished from similar Category 2 species of *Acanthobothrium* that have been described from the Pacific Coast of the Americas (amphi-American species), and from others parts of Pacific Ocean by the measurements given in the Table 1. The total length of the new species (2.9–6.7 mm) is shorter than that of *A. campbelli* (0.99–1.8 mm). The number of proglottids of the new species (10–13) is less than that of *A. annapinkiensis* Carvajal & Goldstein, 1971 (15–26), *A. cimari* Marques, Brooks & Monks, 1995 (14–33), *A. puntarenasense*

Table 1. Comparison of *Acanthobothrium soniae* sp. nov. vs. Category 2 species of the genus from the Pacific Ocean. Abbreviations: No. Number; A Base (handle) length; B Axial prong length; C Abaxial prong length; D Total hook length. Note: the use of "–" without numerical values are measurement ranges that overlap those of *A. soniae* sp. nov.

Species of *Acanthobothrium*	Total length (mm)	No. of proglottids	Diameter of accesory sucker (µm)	Length of cirrus sac (µm)	No. of testes per proglottid	Measurements of hook (µm)			
						A	B	C	D
Pacific coast of the Americas									
A. soniae sp. nov.	2.9–6.7	10–13	45–50	55–90	31–47	43–45	88–105	83–93	125–138
A. annapinkiensis Carvajal & Goldstein, 1971	–	15–26	~ 120	–	–	60–80	180–250	160–240	240–310
A. brachyacanthum Riser, 1955	–	–	–	123–135	–	–	66	57–60	90
A. bullardi Ghoshroy & Caira, 2001	–	–	–	113–175	–	–	–	43–78	–
A. campbelli Marques, Brooks & Monks, 1995	0.99–1.8	3–6	16–38	–	15–23	16–41	–	–	95–120
A. cimari Marques, Brooks & Monks, 1995	–	14–33	–	148–180	–	–	–	57–82	–
A. coquimbensis Carvajal & Jeges, 1980	–	–	16–44	200–400	–	–	–	96–136	–
A. costarricense Marques, Brooks & Monks, 1995	–	–	–	110–236	–	–	–	54–66	–
A. dasi Ghoshroy & Caira, 2001	–	–	–	100–153	–	–	–	50–75	68–125
A. olseni Dailey & Mudry, 1968	–	–	24–34	96–168	–	–	–	–	91–115
A. puntarenasense Marques, Brooks & Monks, 1995	–	23–27	14–15	151–183	–	–	72–82	75–81	107–114
A. rajivi Ghoshroy & Caira, 2001	–	–	–	–	9–13	28–35	63–73	58–68	88–98
A. unilateralis Alexander, 1953	–	–	–	150–160	–	58–64	118	118	173–182
A. vargasi Marques, Brooks & Monks, 1995	–	5–7	22–41	–	22–29	–	–	–	–
Other parts of the Pacific Ocean									
A. guanghaiense Yang, Sun, Zhi, Iwaki, Reyda & Yang, 2016	–	13–28	28–32	95–132	–	–	–	94–124	–
A. masnihae Fyler & Caira, 2006	–	23–43	–	–	6–12	–	69–88	–	99–123
A. popi Fyler, Caira & Jensen, 2009	–	14–20	50–88	108–152	–	–	–	–	–
A. tetabuanense Reyda & Caira, 2006	–	25–36	–	–	6–12	–	–	–	–

(23–27), *A. guanghaiense* Yang, Sun, Zhi, Iwaki, Reyda & Yang, 2016 (13–28), *A. masnihae* Fyler & Caira, 2006 (23–43), *A. popi* Fyler, Caira & Jensen, 2009 (14–20), and *A. tetabuanense* Reyda & Caira, 2006 (25–36), and the number of proglottids of the new species is greater than that of *A. campbelli* (3–6) and *A. vargasi* (5–7). The diameter of the accessory sucker of the new species (45–50) is shorter than that *A. annapinkiensis* (~120) and *A. popi* (50–88), and the diameter of the accessory sucker of the new species is longer that *A. campbelli* (16–38), *A. coquimbensis* Carvajal & Jeges, 1980 (16–44), *A. olseni* Dailey & Mudry, 1968 (24–34), *A. puntarenasense* (14–15), *A. vargasi* (22–41), *A. guanghaiense* (28–32) and *A. popi* (50–88). The length of the

cirrus sac of the new species (55–90) is shorter than that of *A. brachyacanthum* Risen, 1955 (123–135), *A. bullardi* (113–175), *A. cimari* (148–180), *A. coquimbensis* (200–400), *A. costarricense* (100–236), *A. dasi* (100–153), *A. olseni* (96–168), *A. puntarenasense* (151–183), *A. unilateralis* Alexander, 1953 (150–160), *A. guanghaiense* (95–132), and *A. popi* (108–152). The number of testes per proglottid of the new species (31–47) is less than that of *A. campbelli* (15–23), *A. rajivi* (9–13), *A. vargasi* (6–12), and *A. tetabuanense* (6–12). Finally the measurements of the hooks of the 18 species can be found in Table 1.

Acanthobothrium vidali sp. nov.

http://zoobank.org/9D9106EF-7772-4E61-A44F-B2D151782329

Figures 3A–D; 4A–D

Type material. Holotype (CNHE-11134), 7 paratypes (CNHE-11135), 3 paratypes (HWML-139979, 139980, 139981), and 7 paratypes (CHE-P00082).

Other material examined. *Acanthobothrium franus* Marques, Centritto & Stewart, 1997 (CNHE–3140, Costa Rica) paratype; *A. inbiorium* Marques, Centritto & Stewart, 1997 (CNHE–3138, Costa Rica) paratype; *A. obuncus* Marques, Brooks & Barriga, 1997 (CNHE–3032A, 3167B, Ecuador) holotype; *A. soberoni* Ghoshroy & Caira, 2001 (CNHE–4042, México).

Type host. *Narcine entemedor* Jordan & Starks, 1895 (Elasmobranchii: Torpediniformes: Narcinidae).

Type locality. Bahía de Acapulco (Playa Las Hamacas: 16°51'11"N, 99°53'59"W), Guerrero, México.

Site of infection. Spiral intestine.

Quantitative descriptors of parasite populations (Bush et al. 1997). Prevalence = 36.36% (4 of 11 rays were infected); abundance = 1.91 (21 helminths in 11 rays); mean intensity = 5.25 (21 helminths in 4 infected rays).

Etymology. The species is named in honor of Dr Victor Vidal Martínez (Departamento de Recursos del Mar, CINVESTAV-IPN, Merida, Yucatan, Mexico), for his contribution to our knowledge of helminths of fishes from Mexico.

Diagnosis. *Acanthobothrium vidali* sp. nov. is a Category 6 species. This species is large, with a range of 164–214 craspedote proglottids, with a range of 50–76 testes per proglottid, and the arms of ovary unequal in length (asymmetrical). This new species also can be distinguished from similar congeners by total length, number of proglottids, diameter of accessory sucker, the length of the cirrus sac, number of testes per proglottid, and size of the hooks.

Description. [Based on 5 complete worms and 16 partial specimens] Worms 26.5–70.9 mm (n = 5) long, greatest width at level of mature proglottids, euapolytic; 164–214 (n = 5) proglottids per worm. Scolex 880–1,400 (n = 20) long by 680–1,170 (n = 20) wide, with four bothridia; maximum width of scolex at level of posterior margin of anterior loculus (Figs 3A, 4A). Bothridia free posteriorly, tri-locular, 770–1,230

(n = 20) long by 320–570 (n = 20) wide, with anterior muscular pad (Figs 3A, 4A). Muscular pad 250–325 (n = 18) wide, with apical sucker 75–150 (n = 19) and one pair of bipronged hooks at posterior margin (Figs 3A, 4A). Anterior loculus of bothridia 400–650 (n = 20) long; middle loculus 170–310 (n = 20) long; posterior loculus 150–340 (n = 20) long (Figs 3A, 4A); loculus length ratio (anterior:middle:posterior) 1:0.48:0.50. Velum between medial margins of bothridia in dorsal or ventral pairs not seen (Figs 3A, 4A). Hooks bipronged, hollow, with tubercle on proximal surface of axial prong; internal channels of axial and abaxial prongs continuous, smooth, base and anterior part of each hook embedded in musculature of scolex, tips of prongs free (Figs 3A, B, 4A). Bases (handles) of medial and lateral hooks articulate with one another (Figs 3B, 4A). Lateral hook measurements (n = 15): A 140–170, B 200–285, C 140–305, D 360–465; Medial hook measurements (n = 15): A' 100–165, B' 225–300, C' 200–270, D' 300–425. Cephalic peduncle 2.38–9.13 mm (n = 15) long by 0.15–0.23 mm (n = 15) wide, microtriches not seen on the scolex or cephalic peduncle (Figs 3A, 4A, B, C). Proglottids craspedote. Immature proglottids 50–230 (n = 17) long by 240–520 (n = 17) wide, mature proglottids 260–700 (n = 10) long by 300–790 (n = 10) wide (Fig. 3C), terminal proglottids 1,120 (n = 1) long by 480 (n = 1) wide. Genital pore marginal, irregularly alternating, 49%–63% (n = 9) of proglottid length from anterior end in mature proglottids; genital atrium present (Figs 3C, D, 4D). Testes arranged in two to three irregular columns on each side of the uterus, in frontal view testes wider than long in mature proglottids, 50–125 (n = 6) long by 40–50 (n = 6) wide (Fig. 3C). In terminal proglottids, anteriormost testes wider than long and posteriormost testes longer than wide. Total number of testes 50–76 (n = 10), aporal 26–40 (n = 10), poral 23–36 (n = 10), preporal 17–26 (n = 10), postporal 5–11 (n = 10). All testes located anterior to ovarian isthmus. Cirrus sac pyriform, 125–175 (n = 6) long by 30–75 (n = 6) wide in mature proglottids (Figs 3C, D, 4D). Cirrus armed. Vagina anterior to cirrus sac (Figs 3C, D, 4D),walls relatively thick, covered with gland cells. Vagina extending laterally from common genital atrium, following anterior margin of cirrus sac, weakly sinuous posteriorly along medial line of proglottid to oötype (Fig. 3C); vaginal sphincter absent. Seminal receptacle not seen. Ovary in mature proglottids H-shaped in frontal view (Fig. 3C); posterior lobes wider than anterior lobes. Ovarian isthmus approximately 2/3 of the distance from anterior end of ovary. Arms of ovary unequal in length, aporal arm always longer than poral arm (Fig. 3C). Aporal arm 150–260 (n = 8) long, reaching to posterior margin of cirrus sac, poral arm 125–225 (n = 8) long in mature proglottids, not reaching posterior margin of cirrus sac. Mehlis' gland posterior to ovarian isthmus. Vitiellarium follicular, forming lateral bands, extending from near anterior margin of proglottid to near posterior margin of proglottid (Fig. 3C); follicles 15–20 (n = 8) long by 10–15 (n = 8) wide. Uterus thin-walled, saccate, extending from anterior margin of proglottid to near posterior margin of proglottid. Excretory ducts lateral. Gravid proglottids and eggs not seen.

Remarks. There are 14 Category 6 species of *Acanthobothrium* that have been described worldwide. Of these, seven species have been found in the Pacific Ocean, four of which are amphi-American (Table 2). *Acanthobothrium vidali* sp. nov. also is a

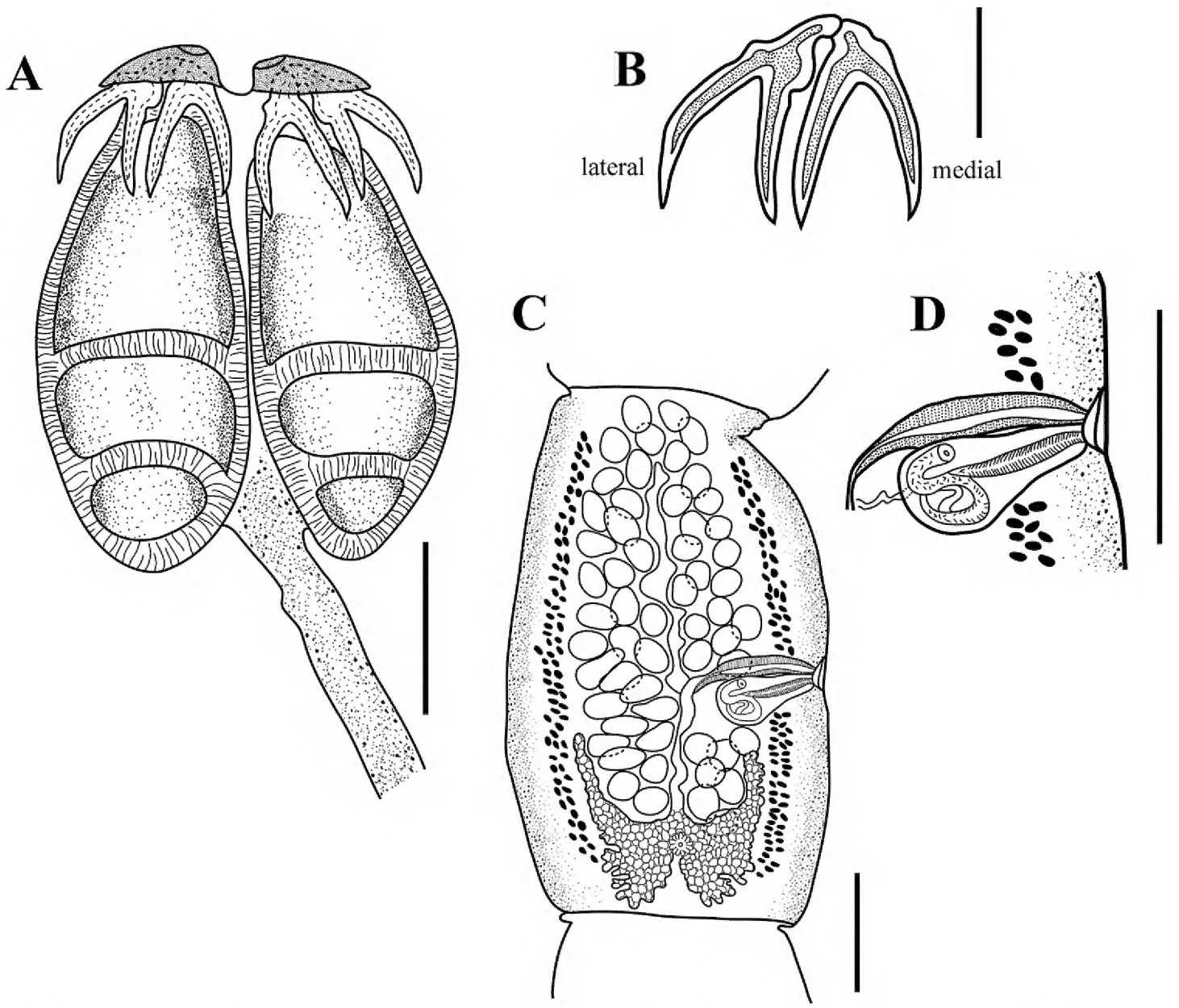

Figure 3. Holotype of *Acanthobothrium vidali* sp. nov. (CNHE-11134). **A** Scolex **B** hooks **C** mature proglottid **D** genitalia. Scale bars: 400 μm (**A**); 200 μm (**B–D**).

Category 6 species, bringing the total number to 15. Category 6 species have a total length >15 mm (the length of *A. vidali* sp. nov. is 26.5–70.9 mm), a strobila made up of > 50 proglottids (*A. vidali* sp. nov. has 164–214 proglottids), number of testes per proglottids ≤ 80 (*A. vidali* sp. nov. has 50–76 testes per proglottid), and the arms of the ovary are asymmetrical.

As presented in Table 2, the new species can be distinguished from similar Category 6 species of *Acanthobothrium* that have been described from the Pacific Coast of the Americas (amphi-American species), and from others parts of Pacific Ocean by the measurementes given in Table 2. The total length of the new species (26.5–70.9 mm) is longer than that of *A. aetiobatidis* (Shipley, 1900) Southwell, 1925. The number of proglottids of the new species (164–214) is greater than that of *A. gonzalesmugaburoi* Severino & Sarmiento, 1979. The diameter of accessory sucker of the new species (75–150) is larger than that of *A. obuncus* (33–48) and *A. soberoni* (40–65). The length of the cirrus sac of the new species (125–175) is shorter than that of *A. obuncus* (258–322), *A. aetiobatidis* (200–250), and *A. rodmani* Fyler, Caira & Jensen, 2009 (190–234). The number

Figure 4. Light microscope photographs of holotype of *Acanthobothrium vidali* sp. nov. (CNHE-11134). **A** Scolex **B** details of bothridium **C** peduncle cephalic **D** genitalia. Scale bars: 400 μm (**A**); 40 μm (**B**);100 μm (**C, D**). Abbreviations: vg vagina; cs cirrus sac; gp genital pore.

Table 2. Comparison of *Acanthobothrium vidali* sp. nov. vs. Category 6 species of the genus from the Pacific Ocean. Abbreviations: No. Number; A Base (handle) length; B Axial prong length; C Abaxial prong length; D Total hook length. Note: the use of "–" without numerical values are measurement ranges that overlap those of *A. vidali* sp. nov.

Species of *Acanthobothrium*	Total length (mm)	No. of proglottids	Diameter of accesory sucker (μm)	Length of cirrus sac (μm)	No. of testes per proglottid	Measurements of hook (μm)			
						A	B	C	D
Pacific coast of the Americas									
A. vidali sp. nov.	26.5–70.9	164–214	75–150	125–175	50–76	140–170	200–285	140–305	360–465
A. gonzalesmugaburoi Severino & Sarmiento, 1979	–	38–63	–	–	–	50–87	67–123	57–120	146–219
A. maculatum Riser, 1955	–	–	–	–	–	72–78	72–78	75–78	135–141
A. obuncus Marques, Brooks & Barriga, 1997	–	–	33–48	258–322	–	66	60–64	63–64	126–130
A. soberoni Ghoshroy & Caira, 2001	–	–	40–65	–	–	45–88	43–90	65–100	86–158
Other parts of the Pacific Ocean									
A. aetiobatidis (Shipley, 1900), Southwell, 1925	15–20	–	–	200–250	23–28	–	120–130	120–130	250–280
A. arlenae Campbell & Beveridge, 2002	–	–	–	–	17–26	–	160–179	–	289–344
A. rodmani Fyler, Caira & Jensen, 2009	–	–	–	190–234	17–26	–	–	–	335–357

Table 3. Comparison of species of *Acanthobothrium* that have been reported from *Narcine entemedor*. Abbreviations: No. Number; A Base (handle) length; B Axial prong length; C Abaxial prong length; D Total hook length. Note: Information taken from the original descriptions and this study.

Species of *Acanthobothrium*	Total length (mm)	No. of proglottids	Diameter of accesory sucker (µm)	Length of cirrus sac (µm)	No. of testes per proglottid	Measurements of hook (µm)			
						A	B	C	D
Reported from *Narcine entemedor*									
A. franus Marques, Centritto & Stewart, 1997*	16.0–40.0	68–141	60–159	102–281	24–59	118–175	245–319	223–322	354–465
A. inbiorium Marques, Centritto & Stewart, 1997	28.0–82.0	156–223	20–75	122–285	44–73	35–50	65–75	50–60	95–120
A. soniae sp. nov.	2.9–6.7	10–13	45–50	55–90	31– 47	43–45	88–105	83–93	125–138
A. vidali sp. nov.	26.5–70.9	164–214	75–150	125–175	50– 76	140–170	200–285	140–305	360–465

of testes per proglottid of the new species (50–76) is greater than that of *A. aetiobatidis* (23–28), *A. arlenae* Campbell & Beverage, 2002 (17–26), and *A. rodmani* (17–26). Finally, the measurements of the hooks of the eight species can be found in Table 2.

The new species is the fourth species of *Acanthobothrium* reported from *N. entemedor*, preceded by *A. franus* and *A. inbiorium* (Category 5 species), and *A. soniae* sp. nov. (Category 2 species) described above. All species have been reported from the Pacific Coast of the Americas. *Acanthobothrium vidali* sp. nov. can be distinguished from these other species by number of proglottids (164–214) is greater than that of *A. franus* (68–141) and *A. soniae* sp. nov. (10–13). The total length of the lateral hook of *A. vidali* sp. nov. (360–465) is longer than that of *A. inbiorium* (95–120 µm). The length of the axial prong of the lateral hook of *A. vidali* sp. nov. (200–285) is longer than that of *A. inbiorium* (65–75 µm) (Table 3).

Discussion

To date, 190 valid species of *Acanthobothrium* have been reported from different regions of the world (Caira et al. 2017; Rodríguez-Ibarra et al. 2018; Franzese and Ivanov 2018). Forty species of *Acanthobothrium* have been described from the Pacific coast of the Americas (eleven species from USA, eleven from México, eight from Costa Rica, four from Ecuador, four from Peru, and three from Chile). With these descriptions of *A. soniae* sp. nov. and *A. vidali* sp. nov., 13 species of *Acanthobothrium* have been reported from the Pacific Coast of Mexico. A list of amphi-American species of *Acanthobothrium* from the Pacific coast, their hosts, and localities is given in Table 4.

Host specificity of most species of *Acanthobothrium* appears to be rather strict (Ivanov, 2005; Vardo-Zalik and Campbell 2011; Franzese and Ivanov 2018). According to the reports of species of the genus (type localities, additional localities, type host, and additional host), 82% of the species of *Acanthobothrium* show strict host specificity. In contrast, 33 of the 190 valid species of *Acanthobothrium* have been reported in more than one species of host (see the reports of Rudolphi 1819; Yoshida

Table 4. Species of *Acanthobothrium* reported from the Pacific Ocean of the Americas (amphi-American species). ‡= Category designation obtained from Ghoshroy and Caira (2001). Category designations not included in Ghoshroy and Caira (2001) were calculated for this study using the original descriptions. Sources were as given by that author or the original descriptions used for this study.

Family / Host species	*Acanthobothrium* species	Type locality	Source	Category designation
Heterodontidae				
Heterodontus francisci (Girard, 1855)	*A. bajaensis* Appy & Dailey, 1973	San Quintin Bay, Baja California, Mexico	Appy and Dailey (1973)	**4‡**
	A. puertecitense Caira & Zahner, 2001	Puertecitos, Gulf of California, Mexico	Caira and Zahner (2001)	**4**
H. mexicanus Taylor & Castro-Aguirre, 1972	*A. santarosaliense* Caira & Zahner, 2001	Santa Rosalia, Gulf of California, Mexico	Caira and Zahner (2001)	**3**
Rhinobatidae				
Pseudobatos productus (Ayres, 1854)	*A. olseni* Dailey & Mudry, 1968	Newport Beach, California, USA	Dailey and Mudry (1968)	**2‡**
	A. rhinobati Alexander, 1953	Santa Monica Harbor, California, USA	Alexander (1953)	**9(5)‡**
	A. robustum Alexander, 1953	Long Beach Harbor, California, USA	Alexander (1953)	**4‡**
Platyrhinidae				
Platyrhinoidis triseriata (Jordan & Gilbert, 1880)	*A. goldsteini* Appy & Dailey, 1973	Seal Beach, California, USA	Appy and Dailey (1973)	**5(9)‡**
Narcinidae				
Diplobatis ommata (Jordan & Gilbert, 1890)	*A. dollyae* Caira & Burge, 2001	Bahía de Los Angeles, Gulf of California, Mexico	Caira and Burge (2001)	**1**
	A. maryanskii Caira & Burge, 2001	Loreto, Gulfo of California, Mexico	Caira and Burge (2001)	**5**
	A. royi Caira & Burge, 2001	Punta Arena, Gulf of California, Mexico	Caira and Burge (2001)	**1**
Narcine entemedor Jordan & Starks, 1895	*A. franus* Marques, Centritto & Stewart, 1997	Cuajiniquil Beach, Gulf of Santa Helena, Guanacaste, Costa Rica	Marques et al. (1997b)	**5(8)‡**
	A. inbiorium Marques, Centritto & Stewart, 1997	Cuajiniquil Beach, Gulf of Santa Helena, Guanacaste, Costa Rica	Marques et al. (1997b)	**5‡**
	A. soniae sp. n.	Playa las Hamacas, Bahía de Acapulco, Guerrero, Mexico	This study	**2‡**
	A. vidali sp. n.	Playa las Hamacas, Bahía de Acapulco, Guerrero, Mexico	This study	**6‡**
Torpedinidae				
Tetronarce californica (Ayres, 1855)	*A. hispidum* Riser, 1955	Monterey Bay, California, USA	Riser (1955)	**5‡**
Rajidae				
Raja stellulata (Gilbert, 1915)	*A. brachyacanthum* Riser, 1955	Monterey Bay, California, USA	Riser (1955)	**2‡**
Zearaja chilensis (Guichenot, 1848)	*A. annapinkiensis* Carvajal & Goldstein, 1971	Anna Pink Hay, Chile	Carvajal-G. and Goldstein (1971)	**2‡**
Arhynchobatidae				
Psammobatis scobina (Philippi, 1857)	*A. psammobati* Carvajal & Goldstein, 1969	South Pacific Ocean, between Papudo and Talcahuano, Chile	Carvajal-G. and Goldstein (1971)	**5‡**
Sympterygia brevicaudata (Cope, 1877)	*A. lusarmientoi* Severino & Verano, 1980	Callao, Lima, Peru	Severino and Verano (1980)	**7**
Gymnuridae				
Gymnura afuerae (Hildebrand, 1946)	*A. atahualpai* Marques, Brooks & Barringa, 1997	Puerto Bolivar, Provincia de El Oro, Ecuador	Marques et al. (1997a)	**1‡**
Dasyatidae				
Hypanus dipterurus (Jordan & Gilbert, 1880)	*A. bullardi* Ghoshroy & Caira, 2001	Bahía de Los Angeles, Gulf of California, Mexico	Ghoshroy and Caira (2001)	**2‡**
	A. dasi Ghoshroy & Caira, 2001	Puertecitos, Gulf of California, Mexico	Ghoshroy and Caira (2001)	**2‡**
	A. rajivi Ghoshroy & Caira, 2001	Puertecitos, Gulf of California, Mexico	Ghoshroy and Caira (2001)	**2‡**
	A. soberoni Ghoshroy & Caira, 2001	Puertecitos, Gulf of California, Mexico	Ghoshroy and Caira (2001)	**6‡**

Family / Host species	*Acanthobothrium* species	Type locality	Source	Category designation
H. longus (Garman, 1880)	*A. cimari* Marques, Brooks & Monks, 1995	Punta Morales, Puntarenas Province, Costa Rica	Marques et al. (1995)	**2‡**
	A. cleofanus Monks, Brooks & Lonce de Leon, 1996	Chamela Bay, Jalisco, Mexico	Monks et al. (1996)	**3‡**
	A. costarricense Marques, Brooks & Monks, 1995	Punta Morales, Puntarenas Province, Costa Rica	Marques et al. (1995)	**2‡**
	A. obuncus Marques, Brooks & Barriga, 1997	Puerto Hualtaco, Provincia de El Oro, Ecuador	Marques et al. (1997a)	**6‡**
	A. puntarenasense Marques, Brooks & Monks, 1995	Punta Morales, Puntarenas Province, Costa Rica	Marques et al. (1995)	**2‡**
	A. vargasi Marques, Brooks & Monks, 1995	Punta Morales, Puntarenas Province, Costa Rica	Marques et al. (1995)	**2‡**
Potamotrygonidae				
Potamotrygon motoro (Müller & Henle, 1841)	*A. peruviense* Reyda, 2008	Madre de Dios River at Boca Manu, Madre de Dios Department, Peru	Reyda (2008)	**1(8)**
Urotrygonidae				
Urobatis halleri (Cooper, 1863)	*A. parviuncinatum* Young, 1954	San Diego Bays, California, USA	Young (1954)	**8‡**
U. tumbesensis (Chirichigno F. & McEachran, 1979)	*A. minusculus* Marques, Brooks & Barriga, 1997	Puerto Hualtaco, Provincia de El Oro, Ecuador	Marques et al. (1997a)	**1‡**
Urotrygon chilensis (Günther, 1872)	*A. campbelli* Marques, Brooks & Monks, 1995	Costa de Pajaros, Puntarenas, Costa Rica	Marques et al. (1995)	**2‡**
Myliobatidae				
Myliobatis californicus Grill, 1865	*A. holorhini* Alexander, 1953	Long Beach Harbor, California, USA	Alexander (1953)	**3‡**
	A. maculatum Riser, 1955	Monterey Bay, California, USA	Rêgo et al. 1968	**6(3)‡**
	A. microcephalum Alexander, 1953	Long Beach Harbor, California, USA	Alexander (1953)	**4‡**
	A. unilateralis Alexander, 1953	Long Beach Harbor, California, USA	Alexander (1953)	**7(2)‡**
M. chilensis Philippi, 1892	*A. coquimbensis* Carvajal & Jeges, 1980	Antofagasta, Chile	Carvajal-G. and Jeges-G. (1980)	**2‡**
M. peruvianus Garman, 1913	*A. gonzalesmugaburoi* Severino & Sarmiento, 1979	Callao, Lima, Peru	Severino and Sarmiento (1979)	**7(6)**
Aetobatidae				
Aetobatus narinari (Euphrasen, 1790)	*A. monksi* Marques, Brooks & Barriga, 1997	Puerto Jelí, Provincia de El Oro, Ecuador	Marques et al. (1997a)	**1‡**
	A. nicoyaense Brooks & McCorquodale, 1995	Punta Morales, Golfo de Nicoya, Costa Rica	Brooks and McCorquodale (1995)	**1‡**
Scombridae (Perciformes)				
Sarda chiliensis (Cuvier, 1832)	*A. chilensis* Rego, Vincednte & Herrera, 1968	Paita, Piúra, Peru	Rêgo et al. (1968)	**3‡**

1917; MacCallum 1921; Léon-Borcéa 1935; Baer 1948; Euzet 1952; Yamaguti 1952; Young 1954; Riser 1955; Rees and Williams 1965; Goldstein 1967; Campbell 1969; Williams 1969; Carvajal-G. and Jeges-G. 1980; Rodriguez and Tantaleán-Vidaurre 1980; Brooks et al. 1981; Mayes and Brooks 1981; Escalante-A. 1986; Tantaleán-Vidaurre 1991; Marques et al. 1997a; Campbell and Beveridge 2002; Friggens and Brown 2005; Lacerda et al. 2008; and, Iannacone et al. 2011).

Prior to de Carvalho and Last (2016), the genus *Narcine* Henle, 1834 was composed of 20 species. To date, those taxa have been divided into two genera; 15 species of *Narcine* (tail length about equal to disc length or width) and 5 species of *Narcinops* de Carvalho & Last, 2016 (tail much longer tan disc length or width), this latter distributed only in

Table 5. Species of *Acanthobothrium* reported in species of *Narcine*. † Data from Ghoshroy and Caira (2001); ‡ Data from Fyler and Caira (2006).

Narcine	Species of	Category	Habitat of host	Type locality	Source
Narcine sp. (Reported as *N. braunii*, synonym of *N. brasiliensis*)	*A. indicum* Subhapradha, 1955	‡ 5	Northern Indian Ocean	Madras Coast, India	Subhapradha (1955)
N. bancroftii (Griffith & Smith, 1834) [reported as *N. brasiliensis* (Olfers, 1831)]	*A. lintoni* Goldstein, Henson & Schlicht, 1968	† 1(8,9,5)	North Carolina to northeastern Brazil	Gulf of Mexico, Texas, USA	Goldstein et al. (1969)
N. brasiliensis	*A. electricolum* Brooks & Mayes, 1978	† 9	Brazil to northern Argentina	Caribbean Sea, near Cartagena, Colombia	Brooks and Mayes (1978)
N. entemedor Jordan & Starks, 1895	*A. franus* Marques, Centritto & Stewart, 1997	† 5(8)	Baja California to northen Peru	Cuajiniquil Beach, Gulf of Santa Helena, Guanacaste, Costa Rica	Marques et al. (1997b)
	A. inbiorium Marques, Centritto & Stewart, 1997	† 5	Baja California to northen Peru	Cuajiniquil Beach, Gulf of Santa Helena, Guanacaste, Costa Rica	Marques et al. (1997b)
	A. soniae sp. nov.	2	Baja California to northen Peru	Playa las Hamacas, Bahía de Acapulco, Guerrero, Mexico	This study
	A. vidali sp. nov.	6	Baja California to northen Peru	Playa las Hamacas, Bahía de Acapulco, Guerrero, Mexico	This study

Australia (Last et al. 2016). No helminths have been reported from the former members of *Narcine* that are now assigned to *Narcinops*. Five valid species of *Acanthobothrium* have been reported worldwide from three species of *Narcine* (Table 5) (Subhapradha, 1955; Goldstein et al. 1969; Brooks and Mayes 1978; Marques et al. 1997b), but no species of *Acanthobothrium* in *Narcine* have been reported from Mexico (Merlo-Serna and García-Prieto 2016). In Mexico, only two species of helminth have been reported previously from *Narcine*: *Anaporrhutum euzeti* Curran, Blend & Overstreet, 2003 and *Nagmia rodmani* Curran, Blend & Overstreet, 2009 (Curran et al. 2003; Curran et al. 2009).

The categorical method suggested by Ghoshroy and Caira (2001) and Fyler and Caira, (2006) was used to facilitate comparisons among the 190 valid species of *Acanthobothrium*. Ghoshroy and Caira (2001) proposed the categories to facilitate comparisons among taxa from the same geographic region. Because of the large number of species worldwide, it is necessary to focus only on those species from the same region that possessing the same combination of characters as the new species; thus, delimiting the comparison between similar species that could be confused with a new species and not comparing each new species to all of the 190 valid species currently described. We agree that comparisons with each species of this expanding group is unnecessary and, as the number of species increases, an exercise in futility. In agreement with previous authors, this categorical method is useful but does not reflect groupings from a rigorous phylogenetic hypothesis (i.e., is phenetic) (Campbell and Beveridge 2002; Ivanov 2005; Reyda and Caira 2006; Twohig et al. 2008; Fyler and Caira 2010; Yang et al. 2016).

Although not all species of the genus have been examined, Franzese and Ivanov (2018) suggest that the pattern of microthiches is quite uniform among species of *Acanthobothrium*; (i.e., all species have filitriches covering most surfaces of the worms,

interspersed with gladiate spinitriches on proximal bothridial surface, scolex proper and the cephalic peduncle). Because of insufficient material, it was not possible to make a study of this species using the SEM, so we cannot provide detailed information on the microtriches.

Acknowledgements

The authors thank all those who made possible the collection and examination of specimens. The authors would like to thank the curator Luis García-Prieto of the CNHE and Scott L Gardner and Gabor Racz (curator and collection manager, respectively) of the HWML, for providing access to laboratories and specimens. Students from the Laboratorio de Morfología Animal, Universidad Autónoma del Estado de Hidalgo, and the Unidad Académica de Ecología Marina, Universidad Autónoma de Guerrero, helped with the collection of specimens. This study was supported by funds from the project "Inventario Ambiental y Establecimiento de Indicadores Regionales de la Red Temática: Calidad Ambiental y Desarrollo Sustentable (PROMEP–SEP)". The Consejo Nacional de Ciencia y Tecnología (CONACYT) provided a scholarship (no. 432427) to FZ–T.

References

Alexander CG (1953) Five new species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from southern California rays. Journal of Parasitology 39: 481–486. https://doi.org/10.2307/3273847

Appy RG, Dailey MD (1973) Two new species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from elasmobranchs of the eastern Pacific. Journal of Parasitology 59: 817–820. https://doi.org/10.2307/3278414

Baer JG (1948) Contributions a l'étude des cestodes de sélaciens I–IV. Bulletin de la Société des Sciences Naturelles de Neuchâtel 71: 63–122.

Blanchard E (1848) Sur l'organisation des vers. Annales des Sciences Naturelles 10: 321–364. https://doi.org/10.5962/bhl.title.51506

Brooks DR, Mayes MA (1978) *Acanthobothrium electricolum* sp. nov. and *A. lintoni* Goldstein, Henson, and Schlicht, 1969 (Cestoda: Tetraphyllidea) from *Narcine brasiliensis* (Olfers) (Chondrichthyes: Torpedinidae) in Colombia. Journal of Parasitology 64: 617–619. https://doi.org/10.2307/3279945

Brooks DR, Mayes MA (1980) Cestodes in four species of euryhaline stingrays from Colombia. Proceedings of the Helminthological Society of Washington 47: 22–29.

Brooks DR, Mayes MA, Thorson TB (1981) Systematic review of cestodes infecting freshwater stingrays (Chondrichthyes: Potamotrygonidae) including four new species from Venezuela. Proceedings of the Helminthological Society of Washington 48: 43–64.

Brooks DR, McCorquodale S (1995) *Acanthobothrium nicoyaense* n. sp. (Eucestoda: Tetraphyllidea: Onchobothriidae) in *Aetobatus narinari* (Euphrasen) (Chondrichthyes: Myliobati-

formes: Myliobatidae) from the Gulf of Nicoya, Costa Rica. Journal of Parasitology 81: 244–246. https://doi.org/10.2307/3283927

Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997) Parasitology meets ecology on its own terms: Margolis et al. revisited. Journal of Parasitology 83: 575–583. https://doi.org/10.2307/3284227

Caira JN (1985) *Calliobothrium evani* sp. nov. (Tetraphyllidea: Onchobothriidae) from the Gulf of California, with a redescription of the hooks of *C. lintoni* and a proposal for onchobothriid hook terminology. Proceedings of the Helminthological Society of Washington 52: 166–174.

Caira JN, Burge AN (2001) Three new species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from the ocellated electric ray, *Diplobatis ommata*, in the Gulf of California, Mexico. Comparative Parasitology 68: 52–65.

Caira JN, Jensen K, Healy CJ (1999) On the phylogenetic relationships among tetraphyllidean, lecanicephalidean and diphyllidean tapeworm genera. Systematic Parasitology 42: 77–151. https://doi.org/10.1023/A:1006192603349

Caira JN, Jensen K, Ivanov VA (2017) Onchoproteocephalidea II. In: Caira JN, Jensen K (Eds) Planetary biodiversity inventory (2008–2017): tapeworms from vertebrate bowels of the earth Special Publication, Natural History Museum, The University of Kansas Lawrence, Kansas, 290–315.

Caira JN, Zahner SD (2001) Two new species of *Acanthobothrium* Beneden, 1849 (Tetraphyllidea: Onchobothriidae) from horn sharks in the Gulf of California, Mexico. Systematic Parasitology 50: 219–229. https://doi.org/10.1023/A:1012241913722

Campbell RA (1969) New species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from Chesapeake Bay, Virginia. Journal of Parasitology 55: 559–570. https://doi.org/10.2307/3277298

Campbell RA, Beveridge I (2002) The genus *Acanthobothrium* (Cestoda: Tetraphyllidea: Onchobothriidae) parastic in Australian elasmobranch fishes. Invertebrate Systematics 16: 273–344. https://doi.org/10.1071/IT01004

Carvajal-G. J, Goldstein RJ (1969) *Acanthobothrium psammobati* n. sp. (Cestoda: Tetraphyllidea: Onchobothriidae) from the skate, *Psammobatis scobina* (Chondrichthyes: Rajiidae) from Chile. Zoologischer Anzeiger 182: 432–435.

Carvajal-G. J, Goldstein RJ (1971) *Acanthobothrium annapinkiensis* n. sp. (Cestoda: Tetraphyllidea: Onchobothriidae) from the skate, *Raja chilensis* (Chondrichthyes: Rajiidae) from Chile. Zoologischer Anzeiger 186: 158–162.

Carvajal-G. J, Jeges-G. J (1980) Cestodos parásitos de *Myliobatis chilensis* Phillipi (Pisces: Myliobatidae), con la descripción de una nueva especie de *Acanthobothrium*. Anales del Centro de Ciencias del Mar y Limnología 7: 51–56.

Chervy L (2009) Unified terminology for cestode microtriches: a proposal from the International Workshops on Cestode Systematics in 2002–2008. Folia Parasitologica 56: 199–230. https://doi.org/10.14411/fp.2009.025

Curran SS, Blend CK, Overstreet RM (2003) *Anaporrhutum euzeti* sp. nov. (Gorgoderidae: Anaporrhutinae) from rays in the Gulf of California, Mexico. In: Combes C, Jourdane J (Eds) Taxonomy, Ecology and Evolution of Metazoan Parasites (Livre homage à Louis Euzet) Tome I. Presses Universitaires de Perpignan, Perpignan, France, 225–234.

Curran SS, Blend CK, Overstreet RM (2009) *Nagmia rodmani* n. sp., *Nagmia cisloi* n. sp., and Probolitrema richiardii (López, 1888) (Gorgoderidae: Anaporrhutinae) from Elasmobranchs in the Gulf of California, Mexico. Comparative Parasitology 76: 6–18. https://doi.org/10.1654/4356.1

Dailey MD, Mudry DR (1968) Two new species of cestodes from California rays. Journal of Parasitology 54: 1141–1143. https://doi.org/10.2307/3276979

de Carvalho MR, Last PR (2016) Chapter 15. Numbfishes, Family Narcinidae. In: Last PR, White WT, de Carvalho MR, Séret B, Stehman MFW, Naylor GJP, McEachran JD (Eds) Rays of the World. Comstock Publishing Associates, a division of Cornell University Press; Clayton South VIC, Australia: CSIRO Publishing, Ithaca, New York, 137–169.

Escalante-A. H (1986) Cestodes de elasmobranquios de la costa peruana. Revista de Ciencias Universidad Nacional Mayor de San Marcos 74: 70–74.

Euzet L (1952) Cestodes tétraphyllides de la côte Atlantique du Maroc et de Mauritanie. (Collection resemblée par R. Ph. Dollfus). Comptes Rendus de la Société des Sciences Naturelles de Maroc 5: 91–96.

Euzet L (1959) Recherches sur les Cestodes Tétraphyllides des Sélaciens de côtes de France. Docteur ès Sciences Naturelles, University of Montpellier, France.

Franzese S, Ivanov VA (2018) Hyperapolytic species of *Acanthobothrium* (Cestoda: Onchoproteocephalidea) from batoids off Argentina. Parasitology International 67: 431–443. https://doi.org/10.1016/j.parint.2018.04.001

Friggens MM, Brown JH (2005) Niche partitioning in the cestode communities of two elasmobranchs. Oikos 108: 76–84. https://doi.org/10.1111/j.0030-1299.2005.13275.x

Froese R, Pauly D (2018) FishBase. Version(02/2018). http://www.fishbase.org

Fyler CA, Caira JN (2006) Five new species of *Acanthobothrium* (Tetraphyllidea: Onchobothriidae) from the freshwater stingray *Himantura chaophraya* (Batoidea : Dasyatidae) in Malaysian Borneo. Journal of Parasitology 92: 105–125. https://doi.org/10.1645/GE-3522.1

Fyler CA, Caira JN (2010) Phylogenetic status of four new species of *Acanthobothrium* (Cestoda : Tetraphyllidea) parasitic on the wedgefish *Rhynchobatus laevis* (Elasmobranchii : Rhynchobatidae): implications for interpreting host associations. Invertebrate Systematics 24: 419–433. https://doi.org/10.1071/IS10034

Ghoshroy S, Caira JN (2001) Four new species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from the whiptail stingray *Dasyatis brevis* in the Gulf of California, Mexico. Journal of Parasitology 87: 354–372. https://doi.org/10.1645/0022-3395(2001)087[0354:FNSOAC]2.0.CO;2

Goldstein RJ (1967) The genus *Acanthobothrium* van Benden, 1849 (Cestoda: Tetraphyllidea). Journal of Parasitology 53: 455–483. https://doi.org/10.2307/3276705

Goldstein RJ, Henson RN, Schlicht FG (1969) *Acanthobothrium lintoni* sp. nov. (Cestoda: Tetraphyllidea) from the electric ray, *Narcine brasiliensis* (Olfers) in the Gulf of Mexico. Zoologischer Anzeiger 181: 435–438.

Iannacone J, Avila-Peltroche J, Rojas-Perea S, Salas-Sierralta M, Neira-Cruzado K, Palomares-Torres R, Valdivia-Alarcón S, Pacheco-Silva A, Benvenutto-Vargas V, Ferrario-Bazalar V (2011) Dinámica poblacional de los parásitos metazoos del pez guitarra del pacífico *Rhinobatos planiceps* (Batoidea: Rajiformes) de la zona costera marina de Lima, Perú. Neotropical Helminthology 5: 265–278.

Ivanov VA (2005) A new species of *Acanthobothrium* (Cestoda: Tetraphyllidea: Onchobothriidae) from the ocellate river stingray, *Potamotrygon motoro* (Chondrichthyes: Potamotrygonidae), in Argentina. Journal of Parasitology 91: 390–396. https://doi.org/10.1645/GE-354R1

Lacerda ACF, Takemoto RM, Pavanelli GC (2008) Digenea, Nematoda, Cestoda, and Acanthocephala, parasites in Potamotrygonidae (Chondrichthyes) from the upper Paraná River floodplain, states of Paraná and Mato Grosso do Sul, Brazil. Check List 4: 115–122. https://doi.org/10.15560/4.2.115

Last PR, White WT, de Carvalho MR, Séret B, Stehman MFW, Naylor GJP, McEachran JD (2016) Rays of the World. Comstock Publishing Associates, a division of Cornell University Press; Clayton South VIC, Australia: CSIRO Publishing, Ithaca, New York, 790 pp.

Léon-Borcéa L (1935) Nouvelle note sur *Acanthobothrium ponticum* L Borcéa (n. sp.). Annales scientifiques de l'Université de Jassy 20: 480–481.

MacCallum GA (1921) Studies in helminthology. Zoopathologica 1: 137–284.

Maleki L, Malek M, Palm HW (2013) Two new species of *Acanthobothrium* (Tetraphyllidea: Onchobothriidae) from *Pastinachus* cf. *sephen* (Myliobatiformes: Dasyatidae) from the Persian Gulf and Gulf of Oman. Folia Parasitologica 60: 448–456. https://doi.org/10.14411/fp.2013.048

Marques F, Brooks DR, Barriga R (1997a) Six species of *Acanthobothrium* (Eucestoda: Tetraphyllidea) in stingrays (Chondrichthyes: Rajiformes: Myliobatoidei) from Ecuador. Journal of Parasitology 83: 475–484. https://doi.org/10.2307/3284414

Marques F, Brooks DR, Monks S (1995) Five new species of *Acanthobothrium* van Beneden, 1849 (Eucestoda: Tetraphyllidea: Onchobothriidae) in stingrays from the Gulf of Nicoya, Costa Rica. Journal of Parasitology 81: 942–951. https://doi.org/10.2307/3284046

Marques F, Centritto R, Stewart AS (1997b) Two new species of *Acanthobothrium* in *Narcine entemedor* (Rajiformes: Narcinidae) from the Northwest Coast of Guanacaste Peninsula, Costa Rica. Journal of Parasitology 83: 927–931. https://doi.org/10.2307/3284291

Merlo-Serna AI, García-Prieto L (2016) A checklist of helminth parasites of Elasmobranchii in Mexico. ZooKeys 563: 73–128. https://doi.org/10.3897/zookeys.563.6067

Mayes MA, Brooks DR (1981) Cestode Parasite of some Venezuelan Stingrays. Proceedings of the Biological Society of Washington 93: 1230–1238.

Monks S, Brooks DR, Pérez-Ponce de León G (1996) A new species of *Acanthobothrium* Van Beneden, 1849 (Eucestoda: Tetraphyllidea: Onchobothriidae) in *Dasyatis longus* Garman (Chondrichthyes: Myliobatiformes: Dasyatididae) from Chamela Bay, Jalisco, Mexico. Journal of Parasitology 82: 484–488. https://doi.org/10.2307/3284090

Monks S, Pulido-Flores G, Lara-Sánchez M (2015a) Distribution extension of *Acanthobothrium cartagenensis* Brooks & Mayes, 1980 (Tetraphyllidea: Onchobothriidae) in *Urobatis jamaicensis* (Cuvier, 1816) (Myliobatiformes: Urotrygonidae) from Quintana Roo, México. Check List 11: 1–3. https://doi.org/10.15560/11.4.1707

Monks S, Zaragoza-Tapia F, Pulido-Flores G, Violante-González J (2015b) A new species of *Serendip* (Cestoda: Tetraphyllidea: Serendipeidae) in *Rhinoptera steindachneri* (Chondrichthyes: Myliobatidae) from the Pacific Coast of Mexico. Comparative Parasitology 82: 262–268. https://doi.org/10.1654/4745.1

Rees G, Williams HH (1965) The functional morphology of the scolex and the genetalia of *Acanthobothrium coronotum* (Rud.) (Cestoda: Tetraphyllidea). Parasitology 55: 617–651.

Rêgo AA, Vicente JJ, Herrera NI (1968) Sôbre dois novos parásitos de peixe da costa do Peru (Cestoda, Tetraphyllidea). Memorias do Instituto Oswaldo Cruz 66: 145–149. https://doi.org/10.1590/S0074-02761968000200002

Reyda FB (2008) Intestinal helminths of freshwater stingrays in southeastern Peru, and a new genus and two new species of cestode. Journal of Parasitology 94: 684–699. https://doi.org/10.1645/GE-1230.1

Reyda FB, Caira JN (2006) Five New Species of *Acanthobothrium* (Cestoda: Tetraphyllidea) from *Himantura uarnacoides* (Myliobatiformes: Dasyatidae) in Malaysian Borneo. Comparative Parasitology 73: 49–71. https://doi.org/10.1654/4194.1

Riser NW (1955) Studies on cestode parasites of sharks and skates. Journal of the Tennessee Academy of Science 30: 265–311.

Rodríguez-Ibarra E, Pulido- Flores G, Violante González J, Monks S (2018) A new species of *Acanthobothrium* (Eucestoda: Onchobothriidae) in *Aetobatus* cf. *narinari* (Myliobatidae) from Campeche, México. Revista Brasileira de Parasitologia Veterinária 27: 66–73. https://doi.org/10.1590/s1984-29612018009

Rodriguez TJ, Tantaleán-Vidaurre M (1980) Estudio sobre helmintos de peces elasmobranquios de la costa Peruana. 1. Nuevos registros de Tetraphyllideos. Boletin Peruano de Parasitologia 2: 71–75.

Rudolphi CA (1819) Entozoorum synopsis, cui accedunt mantissa duplex et indices locupletissimi. Humboldt-Universität, Rücker, 811 pp. https://doi.org/10.5962/bhl.title.9157

Severino LR, Sarmiento BL (1979) Neuva especie del genero *Acanthobothrium* Van Beneden 1849; Cestode: Tetraphyllidea de *Myliobatis peruvianus* Garman 1913. Revista de Ciencias Universidad Nacional Mayor de San Marcos 71: 38–43.

Severino LR, Verano MR (1980) *Acanthobothrium lusarmientoi* n. sp. (Cestoda: Tetraphyllidea: Onchobothriidae) [de] *Psammobatis caudispina* Hildebrand, 1941 (Chondrichtyes: Rajiidae) de Peru. Revista de Ciencias Universidad Nacional Mayor de San Marcos 72: 21–27.

Subhapradha CK (1955) Cestode parasites of fishes of Madras coast. Indian Journal of Helminthology 7: 41–132.

Tantaleán-Vidaurre M (1991) Nuevos helmintos parasitos en peces elasmobranquio de la costa peruana. Boletin de Lima 73: 25–28.

Twohig ME, Caira JN, Fyler CA (2008) Two new cestode species from the dwarf whipray, *Himantura walga* (Batoidea: Dasyatidae), from Borneo, with comments on site and mode of attachment. Journal of Parasitology 94: 1118–1127. https://doi.org/10.1645/GE-1475.1

van Beneden P-J (1849) Notice sur un nouveau genre d'Helminte cestoïde. Bulletin de L'Académie Royale des Sciences, des Lettres et des Beaux-Arts de Belgique 16: 182–193.

van Beneden P-J (1850) Recherches sur la faune littorale de Belgique. Les vers cestoides, considérés sous le rapport physiologique, embryogénique et zooclassique. Mémoires de l'Academie Royale des Sciences, des Lettres et des Beaux-Arts de Belgique 25: 1–204. https://doi.org/10.5962/bhl.title.47103

Vardo-Zalik AM, Campbell RA (2011) Five new species of *Acanthobothrium* van Beneden, 1849 (Cestoda: Tetraphyllidea) in elasmobranchs from the northwest Atlantic and Gulf

of Mexico with the first records from smooth-hound sharks and guitarfish. Zootaxa 2828: 41–64. https://doi.org/10.11646/zootaxa.2838.1.3

Williams HH (1960) A list of parasitic worms, including 22 new records, from marine fishes caught off the British Isles. Annals and Magazine of Natural History 2: 705–715. https://doi.org/10.1080/00222935908655756

Williams HH (1969) The genus *Acanthobothrium* van Beneden 1849 (Cestoda: Tetraphyllidea). Nytt Magasin for Zoologi 17: 1–56.

Yamaguti S (1952) Studies on the helminth fauna of Japan. Part 49. Cestodes of fishes, II. Acta Medica Okayama 8: 1–97.

Yang C, Sun Y, Zhi T, Iwaki T, Reyda FB, Yang T (2016) Two new and one redescribed species of *Acanthobothrium* (Cestoda: Onchoproteocephalidea: Onchobothriidae) from *Dasyatis akajei* (Myliobatiformes: Dasyatidae) in the China Sea. Zootaxa 4169: 286–300. https://doi.org/10.11646/zootaxa.4169.2.3

Yoshida S (1917) Some cestodes from Japanese selachians Including five new species. Parasitology 9: 560–592. https://doi.org/10.1017/S003118200000620X

Young RT (1954) Cestodes of sharks and rays in Southern California. Proceedings of the Helminthological Society of Washington 21: 106–112.